

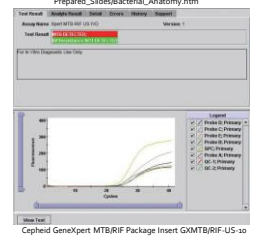
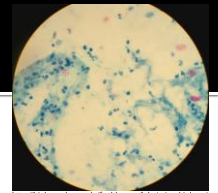
Laboratory Update:

M. tuberculosis (MTB) Characterization, Nucleic Acid Amplification Test (NAAT), & Drug Susceptibility Testing (DST)

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DCLS Microbial Reference Group
Group Manager & Principal Scientist

MTB Identification & Characterization

- Phenotypic Characterization
 - Microscopy
 - Morphology & Biochemical
 - Drug Susceptibility Testing (DST)
- Genotypic Characterization
 - DNA Fingerprinting
 - Spoligotyping
 - RFLP
 - VNTR analysis
 - Probe hybridization
 - Accu Probe
 - Nucleic Acid Amplification Test (NAAT)
 - GenProbe MTD, Cepheid GeneXpert
 - 16 S sequencing



Mycobacteriology Testing: Annual Workload

2012

- Primary Isolation
 - 2963 patient samples
 - 807 individual patients
 - 55 individual patients positive for MTBC
- Reference Culture Identification
 - 616 patients
 - 85 individual patients positive for MTBC
- NAAT Testing (MTD)
 - 74 patients had the GenProbe MTD test performed
 - 30 patients had the MTD detect *M. tuberculosis* DNA

2013 (6 months)

- Primary Isolation
 - 1455 patient samples
 - 387 individual patients
 - 27 individual positive for MTBC
- Reference Culture Identification
 - 371 patients
 - 67 individual patients positive for MTBC
- NAAT Testing (MTD)
 - 26 patients had the GenProbe MTD test performed
 - 5 patients had the MTD detect *M. tuberculosis* DNA

2012 Mycobacteriology Testing: Drug Susceptibility

2012

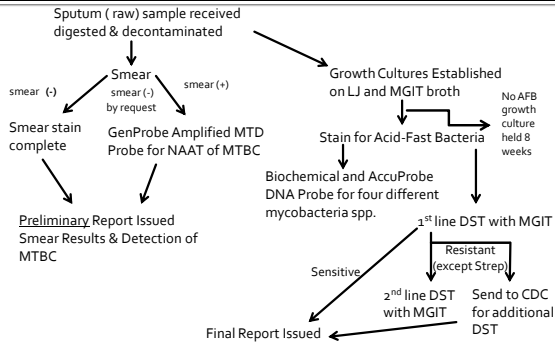
- First Line
 - 157 1st line DST on all initial *M. tuberculosis* isolates
- Second Line
 - 40 2nd line DST on all initial *M. tuberculosis* isolates

2013 (6 months)

- First Line
 - 54 1st line DST on all initial *M. tuberculosis* isolates
- Second Line
 - 12 2nd line DST on all initial *M. tuberculosis* isolates

DST for other *Mycobacteria* spp. available through National Jewish Hospital upon request.

DCLS Current Workflow

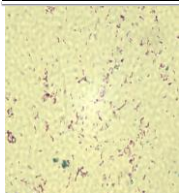


Continuing with the MGIT g60: Primary Isolation, 1st & 2nd Line DST

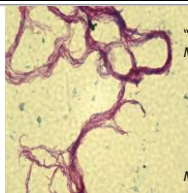
- Continuous incubation at 37°C & monitoring for fluorescence – based on O₂ concentration
- Growth of any organism is detected
 - Mycobacteria, yeast, other bacteria
- Smears prepared from broth
 - Growth determined to be acid-fast



Growth on LJ Slants & Acid Fast Staining



Pleomorphism and branching often seen in *M. avium* complex.



"Cording" typical of *M. tuberculosis*

M. tuberculosis on LJ slants



M. avium on LJ slants



Drug Susceptibility Testing: Performed on MGIT g60

- First line drug testing
 - Isoniazid (INH), Rifampin, Ethambutol, Streptomycin, Pyrazinamide (PZA)
- Results available within 7-12 days after speciation
- Resistant strains - results phoned to submitter
- Second line drug testing
 - Ethionamide, Capreomycin, Ofloxacin, INH at a higher concentration
 - Sent to CDC for additional drug susceptibility testing



GEN-PROBE Amplified MTD Test: DNA Probe for MTBC

- FDA approved the GenProbe Amplified *M. tuberculosis* Direct Test for AFB smear (+) respiratory specimens in 1995 and for smear (-) respiratory specimens in 1999
- Amplified molecular assay detects *M. tuberculosis* directly from sputum samples in less than 3.5 hours

GEN-PROBE Amplified MTD Test: DNA Probe for MTBC

Approved for: Respiratory Specimens

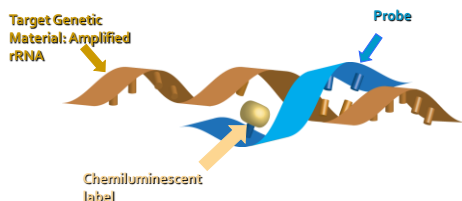
- Testing smear (+) and (-) specimens (NOTE: Smear (-) specimens NOT routinely tested at DCLS)
- **Testing patients who have taken TB medications for LESS than 7 days**
- Patients with high clinical suspicion of TB

NOT Approved for:

- Specimens from patients receiving TB medications in the past 12 months
 - NOT a test of cure
- Testing children or patients unable to produce sputum

GEN-PROBE Amplified MTD Test: DNA Probe for MTBC

- Utilizes a Transcription-Mediated Amplification system (TMA) to detect rRNA



Cepheid GeneXpert MTB /RIF Implementation

- Performed on sputum samples (raw or processed) in ~2h with little hands on time
- In combination with Smear results, detects within 1-2 days of receiving sample:
 - Smear +/-
 - MTBC present
 - Mutation indicative of RIF resistance
- GenProbe Amplified MTD will serve as a backup method

Methodology Comparison

	Gen-Probe Amplified MTD Test [†]	Hains Genotype MTBDR plus Test [‡]	Cepheid GeneXpert MTB/RIF [*]
FDA Approval?	YES	YES	YES
Detection of:	MTBC only	MTBC & Resistance	MTBC & Resistance
Method:	Transcription-Mediated Amplification of rRNA	PCR + DNA-Strip hybridization	Nested real-time PCR
Sample type:	Sputum Sediment and bronchial specimens	Pulmonary specimens & isolates	Raw Sputum or Sputum Sediment
Time-to-Result	2-5-3.5h	5h	2h
Labor Intensive	YES	YES	NO
Detection of MTBC in Smear (+) Sensitivity/Specificity	87.5%/100%	100%/NA	99.7%/98.5%
Detection of MTBC in Smear (-) Sensitivity/Specificity	64.0%/100%	80.3%/98.4%	76.1%/98.8%
RIF Sensitivity/Specificity vs. Convention DST	NA	NA/100% (low sample volume)	94.7%/99.0%
INH Sensitivity/Specificity vs. Convention DST	NA	NA/100% (low sample volume)	NA

[†]GenProbe Amplified MTB users guide IN0044 Rev. P

[‡]Hains Genotype MTBDRplus users guide v2.0 IFU-304A-02. 02/2012

^{*}GeneXpert MTB/RIF users guide 301-1404 Rev. A July 2013

What is the GeneXpert MTB/RIF Assay?

- Nucleic acid amplification test (NAAT)
 - Detects both MTBC and RIF resistance
- Test takes 2h from start to finish

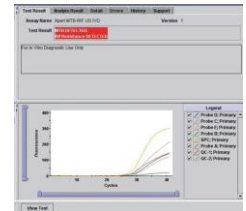
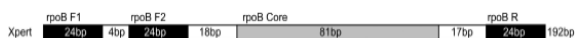


Figure 6. An Example of a MTB DETECTED; RIF Resistance DETECTED Result

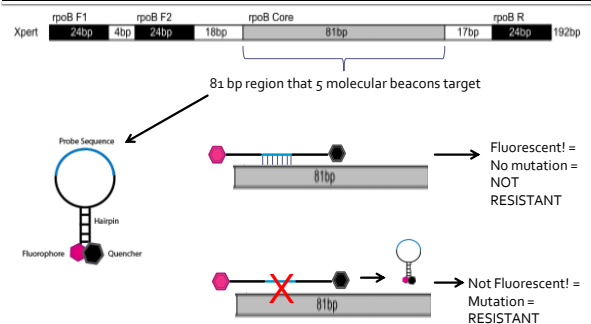
Principle of the GeneXpert Test

- Uses a hemi-nested PCR to amplify the *rpoB* gene in MTB
- Simultaneously probes the PCR amplicon for antibiotic resistance markers.

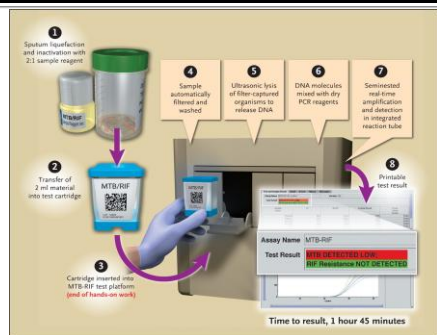


Blakemore, R., et al. 2010. JCM. 48:7. 2495-2501

Principle of the GeneXpert Test part II



GeneXpert MTB/RIF Process



<http://genexpertinfinity.com/>

Sample Requirements

Required Specimen Volume*

Specimen Type	Minimum Volume for One Test	Minimum Total Volume for Test and Retest – See Section 11.2, Retest Procedure
Sputum sediment	0.5 mL	1 mL
Raw sputum	1 mL	2 mL

- Samples should be stored & transported at 2-8°C
- Test is only approved for induced or expectorated sputa
- Samples from patients on antituberculosis drugs for >3 days are NOT acceptable.

*GeneXpert MTB/RIF users guide
301-1404 Rev. A July 2013

Results & Potential Reporting

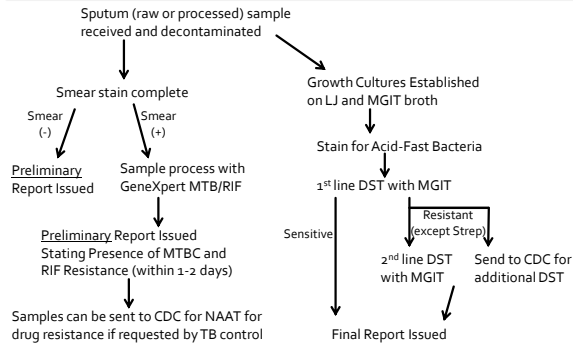
Xpert® MTB/RIF Readout	Interpretation	Report* (Suggested Minimal Language)
MTB DETECTED; RIF Resistance DETECTED	A mutation in the <i>rpoB</i> gene has been detected. A full first and second line drug panel should be conducted.	<i>rpoB</i> mutation detected; likely rifampin resistance; Confirmatory testing in progress OR isolate has been forwarded to a reference laboratory for confirmatory testing.
MTB DETECTED; RIF Resistance NOT DETECTED	A mutation in the <i>rpoB</i> gene has not been detected.	No <i>rpoB</i> mutation detected; likely rifampin susceptible.
MTB DETECTED; RIF Resistance INDETERMINATE	A mutation in the <i>rpoB</i> gene could not be determined due to insufficient signal detection.	Insufficient MTB in the sample to allow determination of <i>rpoB</i> mutation result.
MTB NOT DETECTED	The conserved sequences up- and downstream of the 81bp region were not detected.	MTBC not detected; Confirmatory testing in progress.

APHL Fact Sheet: Sept. 2013

Limitations

- Test has not been evaluated for pediatric patients
- Performance of assay relative to HIV infection status is not known
- Positive MTBC result ≠ viable organisms
- Test does not differentiate between species of MTBC
- Limit of Detection

Proposed DCLS 2014 Workflow



GeneXpert MTB/RIF Initial Presumptive Diagnostic Test

- False positive/negative results are possible
 - Therefore, this test provides preliminary results only
- Smear & MTB/RIF results will be sent as a preliminary report
- Culture and growth based DST will still be conducted
 - MGIT 960 used to confirm
- Final report will be a culmination of all preliminary reports
 - Molecular Detection of Drug Resistance (MDDR) from CDC will be included as an attachment.

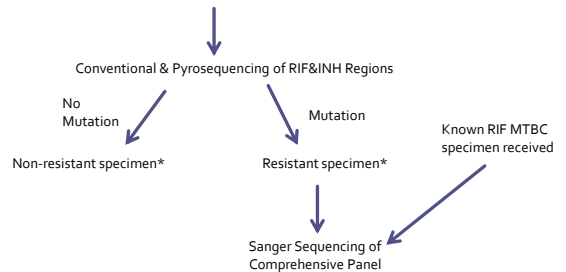
CDC MDDR Assay

- DNA Sequencing to Detect 1st and 2nd line Drug Resistance (2-3 day TAT).

Gene Loci	Associated Antibiotic Resistance
<i>rpoB</i> (81bp region)	Rifampicin
<i>inhA</i> (promoter region)	Isoniazid
<i>katG</i>	Isoniazid
<i>embB</i>	Ethambutol
<i>pncA</i>	Pyrazinamide
<i>gyrA</i>	fluoroquinolones
<i>rrs</i>	Kanamycin, Amikacin, Capreomycin
<i>tlyA</i>	Capreomycin
<i>eis</i> (promoter region)	Kanamycin

CDC MDDR Procedure

NAAT (+) MTBC sputum sample OR Isolate with unknown RIF sensitivity



*Based on NAAT. Confirmation by growth still necessary

Limitations of ALL Molecular Testing

- Gaps in Knowledge
 - What/when do mutations REALLY confer resistance
 - Not all mechanisms of resistance are known
- Limits of Detection
- Absence of mutation does not necessarily mean susceptible
 - Mutation does not necessarily mean resistant
- Conventional detection & DST still required

Conclusions

- DCLS will implement the Cepheid GeneXpert technology in the 1st quarter 2014
- Culture and growth based DST are still the gold standard
- CDC MDDR service can help with early detection of drug resistance. Requests for testing must go through TB Control in collaboration with DCLS
- DCLS and TB Control are available to assist with patient consultations and interpretation of results

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